



Spaceflight-Induced Changes in Microbial Virulence and the Impact to the Host Immune Response

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Introduction

- Over the past 50 years, microorganisms have exhibited unexpected responses relevant to infectious disease when grown in spaceflight and spaceflight analogues, including changes in final cell concentration, biofilm production, stress resistance, antibiotic sensitivity, gene expression, and virulence.
- Seminal studies demonstrated that the foodborne pathogen, *Salmonella enterica* serovar Typhimurium, increased its virulence and pathogenesis-related characteristics, and globally altered its transcriptomic and proteomic profiles, in response to both spaceflight and spaceflight analogue culture¹⁻⁸.



Introduction

- Since those experiments, alterations in the pathogenesis-related characteristics of other pathogens have been documented in response to growth in these environments⁹⁻¹³.
- Notably, a recent study demonstrated an increase in virulence of *Serratia marcescens* cultured during spaceflight, documenting that pathogens other than *Salmonella* can also manifest this response¹⁴.
- However, our overall knowledge of which microorganisms will alter their virulence in response to spaceflight and spaceflight analogue culture, and the underlying mechanisms governing this phenotype, remains very limited.

Experimental Design

- Using bacteria that are cultured in both the spaceflight analogue Rotating Wall Vessel bioreactor (Synthecon) and control conditions, this study will:
 - **Aim 1: Characterize the effect of spaceflight analogue culture on microbial pathogenesis-related stress responses and *in vitro* host-pathogen interactions.** Analysis will include microbial stress responses as well as colonization and viability following pathogen challenge of three-dimensional (3-D) tissue co-culture models containing immune cells.
 - **Aim 2: Characterize the effect of spaceflight analogue culture on the virulence potential of pathogenic microorganisms.** Changes in virulence will be assessed using a mouse model of infection.
 - **Aim 3: Characterize the effect of spaceflight analogue culture on the global molecular genetic responses of pathogenic microorganisms.**

Experimental Design

- We selected obligate and opportunistic pathogens that are medically important and have been or are likely to be found aboard spacecraft, including *Streptococcus pneumoniae*, *Salmonella enterica* serovar Enteritidis, *Pseudomonas aeruginosa*, *Burkholderia cepacia*, and enterohemorrhagic *Escherichia coli* (EHEC).



Progress

- **S. Enteritidis**: Growth kinetics, stress responses, infection of 3-D tissue culture models have been completed. Transcriptomic studies are ongoing.
- **EHEC**: Growth kinetics completed. Stress responses near completion. 3-D tissue culture infections and transcriptomic studies are ongoing.
- **P. aeruginosa**: Growth kinetics and stress responses have been completed. 3-D tissue culture infections and transcriptomic studies are ongoing.
- **B. cepacia**: Growth kinetics and several stress responses have been completed. 3-D tissue culture infections and transcriptomic studies are ongoing.
- **S. pneumoniae**: Growth conditions and media requirements are being optimized.



Significance

- This information will provide critical understanding into the impact of microgravity on potential alterations in microbial virulence and associated infectious disease risk to crew health during spaceflight missions.¹⁵

References

1. Nickerson, C. A., *et al.* (2000) *Infect Immun* **68**(6)
2. Wilson *et al.* (2002) *Appl Environ Microbiol* **68** (11)
3. Wilson *et al.* (2002) *Proc Natl Acad Sci U S A* **99** (21)
4. Wilson, J. W., *et al.* (2007) *Proc Natl Acad Sci U S A* **104**(41)
5. Wilson, J. W., *et al.* (2008) *PLoS One* **3**(12)
6. Barrila, *et al.* (2021) *npj Microgravity* **7**(1)
7. Barrila, *et al.* (2022) *Front Cell Infect Microbiol* 12:705647
8. Franco-Melendez *et al.* (2022) *mSphere*, e0021022
9. Crabbé *et al.* (2010) *Environ Microbiol* **12** (6)
10. Crabbé, A., *et al.* (2011) *Appl Environ Microbiol* **77**(4)
11. Castro, *et al.* (2011) *Appl Environ Microbiol* **77** (18)
12. Crabbé, A., *et al.* (2013) *PLoS One* **8** (12)
13. Yang, J., *et al.* (2016) *npj Microgravity* **2**: 16021
14. Gilbert, R., *et al.* (2020) *npj Microgravity* **6**(4)
15. Nickerson *et al.* (2021) *Nat Microbiol* **7** (4)